REVIEWS AND SYNTHESIS

Poised to prosper? A cross-system comparison of climate change effects on native and non-native species performance

Abstract

Cascade J. B. Sorte, ^{1,*} Ines Ibáñez, ² Dana M. Blumenthal, ³ Nicole A. Molinari, ⁴ Luke P. Miller, ⁵ Edwin D. Grosholz, ⁶ Jeffrey M. Diez, ⁷ Carla M. D'Antonio, ⁸ Julian D. Olden, ⁹ Sierra J. Jones¹⁰ and Jeffrey S. Dukes¹¹ Climate change and biological invasions are primary threats to global biodiversity that may interact in the future. To date, the hypothesis that climate change will favour non-native species has been examined exclusively through local comparisons of single or few species. Here, we take a meta-analytical approach to broadly evaluate whether non-native species are poised to respond more positively than native species to future climatic conditions. We compiled a database of studies in aquatic and terrestrial ecosystems that reported performance measures of non-native (157 species) and co-occurring native species (204 species) under different temperature, CO_2 and precipitation conditions. Our analyses revealed that in terrestrial (primarily plant) systems, native and non-native species responded similarly to environmental changes. By contrast, in aquatic (primarily animal) systems, increases in temperature and CO_2 largely inhibited native species. There was a general trend towards stronger responses among non-native species, including enhanced positive responses to more favourable conditions and stronger negative responses to less favourable conditions. As climate change proceeds, aquatic systems may be particularly vulnerable to invasion. Across systems, there could be a higher risk of invasion at sites becoming more climatically hospitable, whereas sites shifting towards harsher conditions may become more resistant to invasions.

Keywords

Climate change, CO₂, effect size, forecasting, global warming, invasive species, meta-analysis, performance, precipitation, resource utilisation.

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INTRODUCTION

Future climate change may facilitate biological invasions, accentuating its effects on local and regional biodiversity (D'Antonio & Vitousek 1992; Dukes & Mooney 1999; Hellmann et al. 2008; Rahel & Olden 2008; Huang et al. 2011; Diez et al. 2012). Shifts in the magnitude and variability of carbon dioxide (CO₂) levels, temperature and precipitation are altering local conditions, in some cases inhibiting resident species (Walther et al. 2002; Root et al. 2003; Parmesan 2006). These changes may provide colonisation opportunities for non-resident native or non-native species (i.e. species introduced to that location by humans; Richardson et al. 2000; Webber & Scott 2012) that are better suited to the new conditions (Dukes & Mooney 1999; Byers 2002; Thuiller et al. 2007). For example, projected changes in precipitation and temperature could lead to species turnover rates of more than 40% in European plant communities (Thuiller et al. 2005). Although climate change and biological invasions each are altering ecosystem structure and functioning,

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we lack a general, quantitative understanding of how these drivers interact and could synergistically affect ecosystems in the future.

Non-native species may be poised to take advantage of emerging opportunities for colonisation and population growth created by climate change. By definition, non-native species have, given their presence in introduced habitats, already succeeded in colonising new environments. As a result, many non-native species have traits that are useful for coping with environmental change (Dukes & Mooney 1999; Theoharides & Dukes 2007; Vilà et al. 2007), including relatively strong dispersal abilities (Rejmánek & Richardson 1996), minimal reliance on specialised mutualists (van Kleunen et al. 2008), rapid growth rates (Grotkopp et al. 2010), broad environmental tolerances (Willis et al. 2010; Zerebecki & Sorte 2011) and high phenotypic plasticity (Daehler 2003; Davidson et al. 2011). In addition, some climatic changes are increasing resource availability (e.g. increased precipitation and atmospheric CO2) and fluctuations in resource availability (e.g. linked to extreme climatic events; Diez et al. 2012), which could facilitate the establishment and spread of fast-growing species, includ-

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ing many of non-native origin (Davis *et al.* 2000; Daehler 2003; Blumenthal *et al.* 2008; González *et al.* 2010; Dukes *et al.* 2011). Conversely, changes that reduce resource availability, such as decreased precipitation, increased occurrence of drought or CO_2 -driven increases in nitrogen limitation (Daehler 2003; Luo *et al.* 2004), could inhibit non-native species (Bradley *et al.* 2010). Thus, although established non-native species have demonstrated their abilities to persist in new regions, it is not clear whether these species will benefit more than co-occurring native species from changes in climatic conditions.

Relative effects of climate change on native and non-native species are likely to vary widely across ecosystems and taxa. For example, in aquatic systems, elevated CO2 is associated with decreased pH, often inhibiting calcification and growth (Orr et al. 2005). By contrast, elevated CO2 increases carbon availability and enhances water use efficiency for terrestrial plants, increasing growth of most species (Ainsworth & Long 2005) and sometimes strongly favouring nonnative species (Smith et al. 2000; Dukes et al. 2011). Warming may increase growth rates in temperate aquatic and mesic terrestrial ecosystems, thus promoting fast-growing non-native species (Stachowicz et al. 2002; Rahel & Olden 2008; Sorte et al. 2010a); however, in arid and semi-arid ecosystems, increased temperatures may exacerbate drought, potentially favouring drought-tolerant natives (Bradley et al. 2010; Seager & Vecchi 2010). The net effect of climate change on the success of non-native species is likely to depend on both the degree to which environmental alterations inhibit (or promote) native species (Byers 2002) and the availability of both native and non-native species that are better adapted to new conditions (Bradley et al. 2012).

Concerns about how species invasions will interact with climate change have been articulated in several reviews (Dukes & Mooney 1999; Occhipinti-Ambrogi 2007; Thuiller *et al.* 2007; Vilà *et al.* 2007; Hellmann *et al.* 2008; Rahel & Olden 2008; Walther *et al.* 2009; Bradley *et al.* 2010). Until recently, however, there were too few studies comparing native and non-native species responses to predicted climatic conditions to conduct meaningful quantitative syntheses. Here, we provide the first meta-analysis of studies comparing the responses of native and non-native species to elevated CO_2 , warming and changes in precipitation, including studies from terrestrial, marine and freshwater ecosystems. We analysed 132 studies (from 89 publications) that simultaneously quantified performance for both native and non-native species under ambient and altered climatic conditions (Table 1) to address the following questions:

(1) How might climatic changes affect the performance of native and non-native species?

(2) Will predicted climatic conditions differentially favour nonnative species (i.e. do non-natives respond more positively than native species)? (3) How do absolute and relative responses vary by system and environmental driver, as well as by intrinsic attributes (e.g. response type and life stage) and extrinsic factors (e.g. geography and magnitude of climatic change)?

(4) What can the shape of the relationship between performance responses and increasing magnitude of change tell us about which groups of species, under which conditions, exhibit the greatest sensitivity to climate change?

Answering these questions will allow us to assess the combined threat of climate change and biological invasions and to identify drivers that might make particular systems more susceptible to an increase in non-native species. The results of our analyses indicate that altered environmental conditions favoured non-native species in aquatic habitats, but not in terrestrial habitats. However, nonnatives do not appear to be universally poised for increased performance and responded more strongly than native species both to beneficial and detrimental climate changes.

METHODS

We conducted a systematic review of the peer-reviewed literature to support an evidence-based examination of native vs. non-native responses to projected climate change. Systematic reviews follow a strict protocol to maximise transparency and repeatability while minimising bias (Pullin & Knight 2009; Stewart 2010). We applied a set of established guidelines from the ecological sciences for undertaking the steps of a formalised systematic review, which included protocol formation, search strategy, data inclusion, data extraction and analysis (Pullin & Stewart 2006).

Protocol formation and search strategy

We searched ISI Web of Knowledge for topics using a combination of search terms for non-native species, system and environmental driver of climate change, including changes in temperature, CO_2 levels (with aquatic pH) and precipitation (see Appendix S1 in Supporting Information). We also performed targeted searches for cited references as well as publications based on known ongoing global change studies (Terrestrial Carbon (TerraC) Information System 2011).

Data inclusion

In total, we reviewed approximately 60 000 titles and 3000 abstracts to identify articles that met three main criteria: (1) Included at least one native and one non-native species (with origin as identified in the articles themselves or through our own literature search) that

Table 1 Sample sizes for the traditional meta-analysis of performance responses to climate change (with difference-to-mean ratio ES) presented in Figs. 1 and 2. Studies, as defined by independence of controls, were the unit of replication used in the analyses

Climate Change Driver	Articles $(N)/St$	tudies (N)	Native Species Species (N)	(N)/Non-native	Life Form Distribution (% Studies of Plants)	
	Aquatic	Terrestrial	Aquatic	Terrestrial	Aquatic	Terrestrial
+Temperature	13/20	23/26	24/17	68/64	5%	88%
$+CO_2$	5/8	19/23	5/5	58/42	38%	100%
+Precipitation	_	18/23	_	43/26	_	100%
-Precipitation	-	30/35	-	43/37	_	97%

Distribution of life forms is given as the percentage of studies for each driver and system combination focused on plants; the rest of the studies are of animals.

co-occur in the study location, but were not necessarily closely related taxonomically. Non-native species needed to be considered established/naturalised at the study location, but we made no assumptions about species' impacts, (2) contained at least two treatment levels (i.e. ambient and altered conditions) of a particular climate driver and (3) reported a measure of performance that fell within the categories of survival, growth or fecundity.

Data extraction

We identified 89 articles that met our criteria (Appendix S1 and S2), including unpublished data from a dissertation (G. Coffman, Unpublished data) and our own studies (D. Blumenthal & L. Perry, Unpublished data). From these articles, we extracted data for 132 unique studies (including 204 native and 157 non-native species) that were run independently with distinct controls. When necessary, we used digital photo analysis software (e.g. ImageJ; Rasband 2009) to estimate values from published figures. When data were presented for multiple time points in a time series, only the end point (longest duration of the study) was included. When more than two treatment levels were established in a single study, or multiple performance measures were reported, they were all included in our analyses. Performance measure categories included survival (note that mortality estimates were converted to survival rates), growth (biomass, size, cover or photosynthetic rate) and fecundity (number or mass of propagules or reproductive structures). We extracted, when available, mean, sample size and variance for the performance of each species.

Data analysis

We ran two parallel sets of analyses: a traditional meta-analysis and a hierarchical analysis. Within the traditional analysis, we assessed general patterns in responses of native and non-native species to changing climate, and we conducted a mixed-model analysis to investigate effects of potential covariates. In addition, we developed a hierarchical approach to explore the relationship of native and non-native species' responses to increasing magnitudes of climate change.

For both approaches, we calculated the effect size (ES) of each species' response to climate change as the ratio of the difference between treatment and ambient responses to the average of responses across treatment and ambient conditions, or:

$$ES = (response_{treatment} - response_{ambient}) / (\bar{x}_{response})$$

We used this ES instead of the log-response ratio because, while the two metrics are highly correlated (third order polynomial $R^2 = 0.99$), our dataset included a large number of zero values, and the required adjustments for log calculations can influence results (Sweeting *et al.* 2004). For the same reasons, we also used this calculation to estimate magnitude of treatment; thus, the difference between treatment and ambient conditions for the climate driver (i.e. temperature, CO₂ or precipitation) was:

$$MT = (variable_{treatment} - variable_{ambient})/(\bar{x}_{variable})$$

These calculations of effect size and magnitude of treatment allowed us to standardise the treatment conditions and responses across the large variety of studies we worked with, including different climate drivers and different responses (i.e. survival, growth and fecundity). Estimates of both effect size and magnitude of treatment ranged from -2 to 2.

Traditional meta-analysis

We first conducted comparisons to determine the responsiveness to climatic changes across groups and relative differences between native and non-native species. For this analysis, the study was the level of replication, and we pooled ES values for individual species, treatment levels and response types to yield a single ES_{native} and ESnon-native value for each study (i.e. independent comparisons of species' responses, as described above). We then calculated mean effect sizes for the responses of native and non-native species to each climate driver (+temperature, +CO2, +precipitation and -precipitation), both across systems and separately for aquatic (i.e. pooled marine and freshwater) and terrestrial species. We used the jackboot macro in SAS v. 9.2 (SAS Institute 2008) to calculate the bias-corrected bootstrapped 95% confidence interval (based on 999 permutations) for each comparison. Effects on performance of native and non-native species were significant when the bootstrapped confidence intervals did not intersect with zero. To assess whether responses to climate change varied between native and non-native species, we used the methods described above to test for significance of the difference between the responses (i.e. [ES_{non-native}-ES_{native}] calculated separately for each study).

In addition to the study-level analysis above, we conducted a parallel analysis at the individual species level that incorporated the variances in measured performance responses. This analysis comprised a smaller subset of 69% of the studies that reported variances. Further detail on these methods is provided in Appendix S3.

We also used four mixed models to examine whether, at the species level, ES was affected by characteristics of the study treatments, organisms and environments. Mixed models have been used previously for meta-analyses in a variety of research fields, including ecology (Harsch et al. 2009), and offer the flexibility to explore effects of a wide variety of explanatory variables. In all four mixed models, a random effect for the study was used to control for patterns that could be driven only by particular studies. The first model corresponded to the traditional analysis, which addressed whether ES varied for native and non-native species between different study systems (i.e. aquatic and terrestrial) and climate drivers (i.e. + temperature, + CO2, + precipitation and - precipitation). In the second model, magnitude of treatment was added as a covariate to control for differences among studies. The third model included additional study information (treated as fixed effects) that was hypothesised to affect species' responses. These variables were response type (survival, growth and reproduction), habitat [forest, grassland, nongrassland herbaceous, aquatic and other (e.g. desert, shrubland)], geographical location (latitude) and life stage (adult, juvenile and other). The fourth model was used to specifically test for effects of latitude across study systems and climate drivers.

Mixed models to test for effects of additional explanatory variables were fit in a Bayesian framework using OpenBUGS software (Lunn *et al.* 2009) called from R (R Development Core Team 2011) with the package R2OpenBUGS (Sturtz *et al.* 2005), and all model parameters were given non-informative prior distributions. Bayesian meta-analyses using non-informative priors give comparable estimates to traditional methods while offering flexibility to explore more complex models (Mila & Ngugi 2011). Covariates were considered significant if the 95% interval of their coefficients' posterior distributions did not overlap zero. Differences between native and non-native species were assessed by subtracting estimated regression coefficients for natives from those of non-natives, yielding posterior

distributions of the differences between all native and non-native parameters. If the 95% interval of a difference's posterior distribution did not overlap zero, then the responses of native and nonnative species were considered significantly different.

Hierarchical analyses

To examine whether the responses of native and non-native species vary with the magnitude of climate change, we modelled the relationship between effect size and magnitude of the treatment (Osenberg et al. 1997, 1999). To do this, we first divided species according to whether the direction of their responses indicated a detrimental (negative) or beneficial (positive) effect of climate change on performance. We then used absolute values for both variables when estimating effect size as a function of the magnitude of treatment. We initially explored the relationship between the two variables as well as the effect of duration of treatment by fitting several biologically plausible functions to the ES data (e.g. linear, quadratic, logistic). The best-fit relationship (based on lowest Deviance Information Criterion; Spiegelhalter et al. 2000) estimated effect size, ES, as an asymptotic function of magnitude of treatment, with two parameters that describe the maximum effect size and the half saturation constant (see Fig. S4.1 in Appendix S4). These two parameters have useful biological interpretations that can then be compared between native and non-native species: the maximum effect size is an indicator of species' maximum potential responses to climate change, and the half saturation constant indicates how sensitive species' performances are to an increment of change in climatic conditions.

To test for differences between the responses of native and nonnative species under changing conditions, these two parameters were estimated hierarchically. Each parameter's estimates for a particular climate driver (temperature, CO_2 or precipitation) were nested within system (terrestrial or aquatic) and then further nested within an overall estimate for each origin (native or non-native) (Appendix S4; Clark & Gelfand 2006). This hierarchical structure allowed us to test for significant differences between native and non-native species at each level by calculating the differences between each pair of parameters (i.e. *parameter_{native}-parameter_{non-native}*). When 95% confidence intervals around these differences did not include zero, the responses of native and non-native species were considered significantly different.

Finally, we used these parameter values, their means and their variance–covariance matrix to predict effect size as a function of magnitude of treatment at each of the three levels. We used Bayesian methods (Gelman & Hill 2007) for running these hierarchical models in OpenBUGS 1.4 (Thomas *et al.* 2006), and simulations (three chains) were run until convergence of the parameters was ensured (~50 000 iterations). Models were then run for another 25 000 iterations from which posterior parameter values and predicted responses were estimated. Further detail on these methods is provided in Appendix S4.

RESULTS

Our traditional meta-analysis revealed differences in effects of climate change on species performance based on climate drivers and species origins (Fig. 1). For both native and non-native terrestrial species, increased and decreased precipitation led to positive and negative responses respectively. Increased CO_2 benefited non-native spe-



Figure 1 Performance responses of native (black circles) and non-native species (grey triangles) to drivers of climate change (including elevated temperature, CO_2 and precipitation, and decreased precipitation). Effect sizes are given as average ES (difference-to-mean ratio; see *Methods*) for studies of aquatic species (Aq), terrestrial species (Terr) or both (All). Error bars are bias-corrected bootstrapped 95% CIs, and asterisks denote ES's that are different from zero and, thus, significant responses of groups to potential future climatic conditions. Sample sizes are given in Table 1.

cies overall, which was driven by a positive response of terrestrial (primarily plant; Table 1) species. By contrast, aquatic (primarily animal) species, particularly native ones, tended to be negatively affected by increased CO_2 . Temperature effects were non-significant overall and never significant for non-native species. However, there was a positive effect of warming on native terrestrial species and a trend towards a negative effect of warming on native aquatic species. The species-specific results from the variance-weighted analysis always paralleled those from the study-level analysis, with the statistical differences being that the variance-weighted analysis detected significant negative and positive effects of CO_2 enhancement on aquatic and terrestrial natives, respectively, but did not detect significant responses of terrestrial natives under warming or non-natives under increased precipitation (Appendix S3). The mixed-model results presented in Appendix S5 similarly paralleled those presented in Fig. 1.

Results of the paired, within-study analysis indicated that nonnative aquatic species were significantly favoured over native species when temperature was elevated and when CO_2 was increased (Fig. 2). However, in the terrestrial comparisons, no differences were detected between native and non-native species, although nonnatives trended towards a more positive response to increased CO_2 and precipitation and a more negative response than native species to decreased precipitation and increased temperature. The mixed model without additional covariates, an unpaired analysis, gave comparable results: here, non-native species were found to respond significantly more positively than natives under elevated temperatures in aquatic systems and under elevated precipitation in terrestrial systems (Appendix S5).

Of additional factors that we tested via the mixed models, treatment magnitude (i.e. level of environmental change) had significant effects on some response variables: both native and non-native aquatic organisms were more negatively affected in studies with exposure to higher levels of warming (Appendix S5). However,



Figure 2 Differences in effect sizes (ES's; i.e. performance responses) between native and non-native species. Values are mean differences between groups \pm bias-corrected bootstrapped 95% CIs within studies of aquatic species (Aq), terrestrial species (Terr) or both (All). Asterisks denote ES differences between native and non-native species that are significantly greater (non-natives favoured) or less than zero (natives favoured). Sample sizes are given in Table 1.

inclusion of treatment magnitude in the mixed models did not alter the basic estimates of response differences for each origin–driver– system group. No additional factors (including performance response type, habitat, latitude or life stage) had significant effects on native or non-native species responses, although in warmed aquatic systems, the effect of increasing latitude (of the study location) tended to be positive for non-natives but negative for native species (Appendix S5). Overall, there were no significant differences between native and non-native species in how they responded to these covariates (Appendix S5).

In our expanded analysis of the relationship of species' performance responses to magnitude of environmental change, we found that non-native species had higher parameter values (i.e. were more responsive to changing climatic conditions) in all comparisons of the maximum effect size parameter (Fig. 3, Appendix S6). However, in all but two cases, native species were more responsive to increasing treatment magnitude (i.e. had a lower half saturation constant) than non-native species (Fig. 3, Appendix S6). The groups with the maximum potential performance responses to climate change (i.e. largest estimates of maximum effect size) were, for species responding positively, terrestrial and aquatic non-natives under increased temperature and, for species responding negatively, aquatic nonnative species under increased temperature and CO2. The most responsive groups (i.e. groups with the smallest values for the half saturation constant) were all terrestrial species responding positively to precipitation and native aquatic species responding negatively to increased temperature and CO2. The only statistically significant differences between native and non-native species (parameternativeparameternon-native; Fig. 3) were both maximum effect size and half saturation constant for aquatic species responding negatively to temperature increase (Fig. 3, Appendix S6). Overall, although nonsignificant, our predictive curves of effect size as a function of magnitude of treatment suggested that non-native species tended to

respond more strongly both in improved conditions when performance increased, as well as in more stressful conditions when performance decreased (Fig. 4 for overall curves, Appendix S7 for system and driver by system curves).

DISCUSSION

To support proactive ecosystem management in a rapidly changing environment, it is important to understand how ongoing climatic changes are likely to interact with biological invasions. Globally, both factors have been recognised as major drivers of biodiversity loss, and 'interactions among the causes of biodiversity change... represent one of the largest uncertainties in projections of future biodiversity change' (Sala *et al.* 2000). The results of our meta-analysis indicate that absolute and relative responses of native and nonnative species to climatic shifts depend upon changing temperature and the type and direction of altered resource availability. Nonnative species are poised to outperform native species in aquatic ecosystems whereas responses in terrestrial systems are less consistent.

Effects of changing climate on species performance

Our meta-analysis uncovered largely parallel responses of native and non-native species to climate change when resources were either enhanced or became more limiting. For terrestrial species, of which studies of plants composed the majority of data (other studies were of arthropods), our results highlight a pattern of increased performance in response to elevated CO2 and precipitation, but decreased performance at reduced levels of precipitation. The strong responses of terrestrial species to precipitation are consistent with results from a meta-analysis of ecosystem-level responses to changing water availability (Wu et al. 2011). In addition, our finding of a significant increase in performance of nonnative (but not native) terrestrial species under enhanced CO2 is consistent with previous work showing stronger non-native species responses to CO2 enrichment in some studies (e.g. Smith et al. 2000; Belote et al. 2004). Elevated temperature also led to increased plant performance, although only significantly for native species. Responses to warming can also be related - although indirectly - to resource availability: whereas plants in cold-limited and wet climates may typically benefit from warming, those in waterlimited conditions may not (Hoeppner & Dukes 2012). A post hoc comparison indicated that effects of warming (for both native and non-native terrestrial species) tended to be negative in arid, but positive in non-arid, ecosystems; however, we were limited in assessing this potential context-dependency by the small number of studies conducted under relatively dry conditions (i.e. 5 of 26 terrestrial studies).

The negative responses of aquatic species – particularly natives – to changing environmental conditions may be related to resource availability or increased metabolic costs. In aquatic systems, increased dissolved CO_2 is associated with a decrease in pH and changes in water chemistry that make shell formation more difficult and costly (Orr *et al.* 2005). Increased temperature generally leads to increased metabolic rates for both aquatic and terrestrial organisms, particularly ectotherms, which represent all of the species included in these studies. Increased temperature also leads to a decrease in dissolved oxygen in aquatic systems, which then fur-



Negative responses to treatment - Detrimental effects of climate change

Figure 3 Responsiveness to treatment magnitude (i.e. magnitude of climatic change) of native (black circles) and non-native species (grey triangles) given as posterior mean values (and 95% CIs) for the parameters of the hierarchical analyses. The maximum effect size is indicative of the maximum change in performance with climate change, whereas a lower half saturation constant indicates greater sensitivity to increasing magnitude. Parameters were estimated at the overall, system, and driver-within-system levels separately for negative and positive responses to altered climatic conditions for terrestrial (T) and aquatic (A) species. Asterisks denote statistically significant differences between natives and non-native species.

ther lowers the tolerance of aquatic animals to warming (Pörtner & Knust 2007). Changes in resource availability could have, then, driven a number of the performance responses that we detected across native and non-native species in both aquatic and terrestrial systems.

Will non-native species be favoured under climate change?

We found that performance of aquatic non-native species decreased less than that of co-occurring native species in potential future climatic conditions, whereas we found only weak evidence for differential responses in terrestrial ecosystems. The lack of a strong and consistent origin-related response of terrestrial species to climatic factors of global change contrasts with results found, for example, in a meta-analysis of responses to eutrophication: nutrient enrichment consistently favoured non-native plants and invertebrates over their native counterparts (González *et al.* 2010). Recognising distinctions between study designs is important for interpreting differing results across analyses of performance responses to climate change.

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In this study, we quantified how predicted climatic conditions changed performance of native and non-native species relative to current ambient or average conditions, rather than comparing absolute performance differences between native and non-native species (e.g. González et al. 2010). Therefore, our findings for terrestrial species suggest that responses to climate change will not differ between native and non-native species; however, if the current trend is for non-natives to outperform native species, then there is no climatebased reason for this to change in the future. Results from a metaanalysis of performance-related traits in plants yield support for the hypothesis that non-natives outperform native species under current climatic conditions in some settings (van Kleunen et al. 2010). Furthermore, in a post boc analysis of the performance differences between native and non-native plant species in our study (using the effect size ES for the ambient responsenon-native vs. responsenative), we detected a slight non-native performance advantage (0.15 ± 0.08 SE; one-sample t-test t = 1.880, d.f. = 93, P = 0.063). Thus, in terrestrial plant systems, the lack of differential responses to altered conditions would suggest that non-native species are likely to at



Figure 4 Observed (symbols) and predicted effect size (mean middle lines, and 95% PI lower and upper lines) as a function of magnitude of climate-change treatment. Responses were analysed separately for (a) negative and (b) positive responses of native (black circles and solid lines) and non-native species (grey triangles and dashed lines).

least retain any prior advantage over native species as the climate changes.

In aquatic ecosystems, our results suggest that non-native species are favoured under environmental change relative to native species. Non-native species were less negatively affected by increases in both temperature and CO₂ than co-occurring native species. This dichotomy of non-native performance advantages under climate change in aquatic but not terrestrial systems is an interesting finding, but has an important caveat: we were unable to distinguish among differences between native and non-native species that are innate to system (i.e. aquatic or terrestrial) or to life form (i.e. plant or animal). This is because, although we conducted our analyses hierarchically by climate driver (i.e. temperature, CO2 and precipitation) and system, we necessarily relied on a subset of organisms that are amenable to experimentation and observation, and thus the focus of past study. As a result, there was a disproportionate representation of animals (particularly invertebrates) in aquatic studies (although less so in the CO₂ analyses) and plants in terrestrial studies (Table 1; Appendix S2). For example, although responses of aquatic species to increased CO2 were, on balance, negative, this was driven by the negative animal responses: non-native and native aquatic primary producers responded positively in 3/3 and 2/3 of studies respectively.

Thus, particularly in aquatic animal systems exposed to warming or acidification, non-native species appear to be at a performance advantage relative to co-occurring native species. Mechanisms for this pattern may include differences between native and non-native species in environmental conditions at their geographical origins and their respective physiological tolerances (e.g. see Deutsch *et al.* 2008). For the species compared in several of these studies, compilations of experimental results indicate that the non-natives can tolerate higher – and a broader range of – temperatures [e.g. for the Mediterranean mussel (Lockwood & Somero 2011; Somero 2012) and an assemblage of non-native invertebrates (Zerebecki & Sorte 2011)]. Therefore, warming conditions can sometimes become more physiologically optimal for particular species (e.g. Witte *et al.* 2010). Furthermore, all of the aquatic experiments were conducted in temperate habitats, whereas the majority of the aquatic non-natives originated in warmer locations (e.g. the Mediterranean or northwestern Pacific), indicating that the non-native advantage may derive from a long history of adaptation to higher temperatures. The importance of geography is also illustrated, to some degree, by the mixed-model results, which suggest that native species inhabiting cooler (higher latitude) locations are most negatively affected at increased temperature, whereas the non-natives in these locations are poised for more positive performance responses to warming. Unlike the warming comparisons, there are few studies available to assess physiological mechanisms that may explain differential CO2 or pH tolerances between native and non-native species. It is also interesting to note that patterns of thermal tolerance and latitudinal variation did not lead to differential native vs. non-native performance responses for terrestrial plants (Fig. 2).

The uneven taxonomic and geographical distribution of studies in our database highlights the need for additional study of the responses of native and non-native terrestrial animals and aquatic primary producers to climate change, especially in non-temperate habitats. For example, a recent literature review revealed that only a small fraction of non-native species have been well studied (only 49 of 892 species were the subject of 10 or more studies), and only in a subset of geographical regions, with Africa and Asia understudied (Pyšek *et al.* 2008). Although we compiled data from a relatively large number of studies for this meta-analysis, our sample sizes were limited for particular combinations of systems and drivers (Table 1). The fact that most non-significant trends matched predictions for differential native vs. non-native responses (Dukes & Mooney 1999; Rahel & Olden 2008; Bradley *et al.* 2010) suggests that stronger patterns could emerge as more data become available.

Shape and sensitivity of responses to climate change

Beyond the absolute and relative directions of their performance responses, our analyses indicated that non-native species tended to respond more strongly than native species either when conditions became more suitable (increased survival, growth, fecundity, etc.) or when conditions became more stressful (i.e. increased mortality or stunted growth) (Fig. 3). These patterns appear characteristic of opportunistic species that are able to quickly capitalise on increased resources such as enhanced precipitation or elevated CO2 but, at the same time, may not perform as effectively through stressful periods (Davis et al. 2000; Blumenthal 2006). For growth and reproduction, greater responsiveness of non-native species is also consistent with non-native species having higher phenotypic plasticity and incurring increased cost under resource limitation - as compared with native species (Daehler 2003; Davidson et al. 2011). Across our analyses, however, we observed large variability in responses within groups, which led to large variation in predictive curves of performance responses as a function of magnitude of climate change (Fig. 4; Appendix S7). Given these high levels of variability, statistically significant differences were limited to a single comparison: aquatic species responding negatively to warming. In this case, performance of native species was more responsive to the magnitude of temperature increase, but their decreased performance saturated at a lower level (i.e. relatively less impaired), meaning that aquatic non-natives susceptible to warming had a greater scope for responding negatively to warming.

Describing the relationship of performance to magnitude of climate change allows us to project the relative trajectories of native and non-native species under future climatic conditions. Thus, based on our results for aquatic species that were negatively affected by warming, we might predict non-native species to have an initial advantage, given that performance of native species declined most under relatively moderate changes in climate. However non-natives would sustain greater effects on performance, given their greater response scope as temperatures become increasingly stressful. Furthermore, estimating the slopes of the response curves could allow us to predict relative effects of severely altered climatic conditions outside of the range of climates examined in previous experiments and observations. In summary, given sufficient data, the metrics estimated using this hierarchical approach - sensitivity to magnitude of change and maximum responsiveness - could help us identify ecological thresholds and forecast future ecosystem compositions.

CONCLUSIONS

Our systematic review revealed that in aquatic systems, non-native animal species have a strong performance advantage associated with increases in temperatures and CO2 levels. We also identified weaker trends towards similar patterns with increases in CO2 and precipitation among terrestrial species. Increasing the disparity in performance between native and non-native species is likely to exacerbate the effects of climate change on community- and ecosystem-level processes, particularly when such non-natives negatively impact resident species. Given our focus on performance measures such as demographic rates (i.e. survival and reproduction) and biomass, components that have the potential to affect abundance, range size and per capita effects, we might speculate that impacts of aquatic non-native species could be enhanced under elevated temperature and CO2 (Parker et al. 1999). Although, in aquatic systems, negative impacts of non-native species have been most often demonstrated (e.g. Williams & Smith 2007; Sorte et al. 2010b), positive impacts could also increase under climate change, and replacement of declining natives might sometimes prove beneficial at the community or ecosystem level (e.g. Crooks 1998). Thus, greater focus on integrating performance measures with an understanding of non-native species' impacts, especially with climate change, is needed for predicting higher level changes under future climatic conditions. In conclusion, we found that non-native species capitalised on increased resources with environmental change, but they were also negatively affected when conditions became less suitable, and that strong differential effects of climate change on native and non-native species are more likely to be observed among aquatic animals than among terrestrial plants.

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STATEMENT OF AUTHORSHIP

CJBS, II and DMB designed the study and database; all authors contributed to the database; CJBS, II, LPM and JMD performed analyses; CJBS, II and DMB led the writing; and all authors contributed to the writing.

REFERENCES

- Ainsworth, E.A. & Long, S.P. (2005). What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. New Phytol., 165, 351–372.
- Belote, R.T., Weltzin, J.F. & Norby, R.J. (2004). Response of an understory plant community to elevated [CO₂] depends on differential responses of dominant invasive species and is mediated by soil water availability. *New Phytol.*, 161, 827–835.
- Blumenthal, D. (2006). Interactions between resource availability and enemy release in plant invasion. *Ecol. Lett.*, 9, 887–895.
- Blumenthal, D., Chimner, R.A., Welker, J.M. & Morgan, J.A. (2008). Increased snow facilitates plant invasion in mixed grass prairie. *New Phytol.*, 179, 440– 448.
- Bradley, B., Wilcove, D. & Oppenheimer, M. (2010). Climate change increases risk of plant invasion in the Eastern United States. *Biol. Invasions*, 12, 1855–1872.
- Bradley, B.A., Blumenthal, D.M., Early, R., Grosholz, E.D., Lawler, J.J., Miller, L.P. *et al.* (2012). Global change, global trade, and the next wave of plant invasions. *Front. Ecol. Environ.*, 10, 20–28.
- Byers, J.E. (2002). Impact of non-indigenous species on natives enhanced by anthropogenic alteration of selection regimes. *Oikos*, 97, 449-458.
- Clark, J.S. & Gelfand, A.E. (2006). A future for models and data in ecology. *Trends Ecol. Evol.*, 21, 375–380.
- Crooks, J.A. (1998). Habitat alteration and community-level effects of an exotic mussel. *Musculista senhousia Mar. Ecol. Progr. Ser.*, 162, 137–152.
- Daehler, C.C. (2003). Performance comparisons of co-occurring native and alien invasive plants: Implications for conservation and restoration. Annu. Rev. Ecol. Evol. Syst., 34, 183–211.

- D'Antonio, C.M. & Vitousek, P.M. (1992). Biological invasions by exotic grasses, the grass fire cycle, and global change. *Annu. Rev. Ecol. Syst.*, 23, 63–87.
- Davidson, A.M., Jennions, M. & Nicotra, A.B. (2011). Do invasive species show higher phenotypic plasticity than native species and, if so, is it adaptive? A meta-analysis. *Ecol. Lett.*, 14, 419–431.
- Davis, M.A., Grime, J.P. & Thompson, K. (2000). Fluctuating resources in plant communities: a general theory of invasibility. J. Ecol., 88, 528–534.
- Deutsch, C.A., Tewksbury, J.J., Huey, R.B., Sheldon, K.S., Ghalambor, C.K., Haak, D.C. et al. (2008). Impacts of climate warming on terrestrial ectotherms across latitude. Proc. Natl Acad. Sci. USA, 105, 6668–6672.
- Diez, J.M., D'Antonio, C.M., Dukes, J.S., Grosholz, E.D., Olden, J.D., Sorte, C. J.B. *et al.* (2012). Will extreme climatic events facilitate biological invasions? *Front. Ecol. Environ.*, 10, 249–257.
- Dukes, J.S. & Mooney, H.A. (1999). Does global change increase the success of biological invaders?. *Trends Ecol. Evol.*, 14, 135–139.
- Dukes, J.S., Chiariello, N.R., Loarie, S.R. & Field, C.B. (2011). Strong response of an invasive plant species (*Centaurea solstitialis* L.) to global environmental changes. *Ecol. Appl.*, 21, 1887–1894.
- Gelman, A. & Hill, J. (2007). Data analysis using Regression and multilevel/bierarchical models. Cambridge University Press, New York, NY.
- González, A.L., Kominoski, J.S., Danger, M., Ishida, S., Iwai, N. & Rubach, A. (2010). Can ecological stoichiometry help explain patterns of biological invasions? *Oikas*, 119, 779–790.
- Grotkopp, E., Erskine-Ogden, J. & Rejmánek, M. (2010). Assessing potential invasiveness of woody horticultural plant species using seedling growth rate traits. J. Appl. Ecol., 47, 1320–1328.
- Harsch, M.A., Hulme, P.E., McGlone, M.S. & Duncan, R.P. (2009). Are treelines advancing? A global meta-analysis of treeline response to climate warming. *Ecol. Lett.*, 12, 1040–1049.
- Hellmann, J.J., Byers, J.E., Bierwagen, B.G. & Dukes, J.S. (2008). Five potential consequences of climate change for invasive species. *Conserv. Biol.*, 22, 534– 543.
- Hoeppner, S.S. & Dukes, J.S. (2012). Interactive responses of old-field plant growth and composition to warming and precipitation. *Glob. Change Biol.*, 18, 1754–1768.
- Huang, D., Haack, R.A. & Zhang, R. (2011). Does global warming increase establishment rates of invasive alien species? A centurial time series analysis. *PLoS ONE*, 6, e24733.
- van Kleunen, M., Manning, J.C., Pasqualetto, V. & Johnson, S.D. (2008). Phylogenetically independent associations between autonomous selffertilization & plant invasiveness. *Am. Nat.*, 171, 195–201.
- van Kleunen, M., Weber, E. & Fischer, M. (2010). A meta-analysis of trait differences between invasive and non-invasive plant species. *Ecol. Lett.*, 13, 235–245.
- Lockwood, B.L. & Somero, G.N. (2011). Invasive and native blue mussels (genus *Mytilus*) on the California coast: the role of physiology in a biological invasion. *J. Exp. Mar. Biol. Ecol.*, 400, 167–174.
- Lunn, D., Spiegelhalter, D., Thomas, A. & Best, N. (2009). The BUGS project: Evolution, critique, and future directions. *Stat. Med.*, 28, 3049–3067.
- Luo, Y., Su, B., Currie, W.S., Dukes, J.S., Finzi, A.C., Hartwig, U. et al. (2004). Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. *Bioscience*, 54, 731–739.
- Mila, A.L. & Ngugi, H.K. (2011). A Bayesian Approach to Meta-Analysis of Plant Pathology Studies. *Phytopathology*, 101, 42–51.
- Occhipinti-Ambrogi, A. (2007). Global change and marine communities: Alien species and climate change. *Mar. Pollut. Bull.*, 55, 342–352.
- Orr, J.C., Fabry, V.J., Aumont, O., Bopp, L., Doney, S.C., Feely, R.A. *et al.* (2005). Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature*, 437, 681–686.
- Osenberg, C.W., Sarnelle, O. & Cooper, S.D. (1997). Effect size in ecological experiments: the application of biological models in meta-analysis. *Am. Nat.*, 150, 798–812.
- Osenberg, C.W., Sarnelle, O., Cooper, S.D. & Holt, R.D. (1999). Resolving ecological questions through meta-analysis: Goals, metrics, and models. *Ecology*, 80, 1105–1117.

- Parker, I.M., Simberloff, D., Lonsdale, W.M., Goodell, K., Wonham, M., Kareiva, P.M. *et al.* (1999). Impact: Toward a framework for understanding the ecological effects of invaders. *Biol. Invasions*, 1, 3–19.
- Parmesan, C. (2006). Ecological and evolutionary responses to recent climate change. Annu. Rev. Ecol. Evol. Syst., 37, 637–669.
- Pörtner, H.O. & Knust, R. (2007). Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science*, 315, 95–97.
- Pullin, A.S. & Knight, T.M. (2009). Doing more good than harm- Building an evidence-base for conservation and environmental management. *Biol. Conserv.*, 142, 931–934.
- Pullin, A.S. & Stewart, G.B. (2006). Guidelines for systematic review in conservation and environmental management. *Conserv. Biol.*, 20, 1647–1656.
- Pyšek, P., Richardson, D.M., Pergl, J., Jarosík, V., Sixtová, Z. & Weber, E. (2008). Geographical and taxonomic biases in invasion ecology. *Trends Ecol. Evol.*, 23, 237–244.
- R Development Core Team (2011). R: A language and environment for statistical computing, R Foundation for Statistical Computing, Vienna, Austria. Available at: http://www.R-project.org/.
- Rahel, F.J. & Olden, J.D. (2008). Assessing the effects of climate change on aquatic invasive species. *Conserv. Biol.*, 22, 521–533.
- Rasband, W.S. (2009). ImageJ. U. S. National Institutes of Health, Bethesda. Available at: http://rsb.info.nih.gov/ij/. Accessed 22 April 2011.
- Rejmánek, M. & Richardson, D.M. (1996). What attributes make some plant species more invasive? *Ecology*, 77, 1655–1661.
- Richardson, D.M., Pyŝek, P., Rejmánek, M., Barbour, M.G., Panetta, F.D. & West, C.J. (2000). Naturalization and invasion of alien plants: concepts and definitions. *Divers. Distrib.*, 6, 93–107.
- Root, T.L., Price, J.T., Hall, K.R., Schneider, S.H., Rosenzweig, C. & Pounds, J. A. (2003). Fingerprints of global warming on wild animals and plants. *Nature*, 421, 57–60.
- Sala, O.E., Chapin, F.S. III, Armesto, J.J., Berlow, E., Bloomfield, J., Dirzo, R. et al. (2000). Global biodiversity scenarios for the year 2100. Science, 287, 1770–1774.
- SAS Institute. (2008). SAS Version 9.2. SAS Institute, Cary, NC.
- Seager, R. & Vecchi, G.A. (2010). Greenhouse warming and the 21st century hydroclimate of southwestern North America. *Proc. Natl Acad. Sci. USA*, 107, 21277–21282.
- Smith, S.D., Huxman, T.E., Zitzer, S.F., Charlet, T.N., Housman, D.C., Coleman, J.S. *et al.* (2000). Elevated CO₂ increases productivity and invasive species success in an arid ecosystem. *Nature*, 408, 79–81.
- Somero, G.N. (2012). The physiology of global change: linking patterns to mechanisms. Ann. Rev. Mar. Sci., 4, 39–61.
- Sorte, C.J.B., Williams, S.L. & Zerebecki, R.A. (2010a). Ocean warming increases threat of invasive species in a marine fouling community. *Ecology*, 91, 2198– 2204.
- Sorte, C.J.B., Williams, S.L. & Carlton, J.T. (2010b). Marine range shifts and species introductions: comparative spread rates and community impacts. *Glob. Ecol. Biogeogr.*, 19, 303–316.
- Spiegelhalter, D.J., Best, N., Carlin, B.P. & Linde, A.V.D. (2000). Bayesian measures of model complexity and fit. J. Royal Stat. Soc. B, 64, 583–639.
- Stachowicz, J.J., Terwin, J.R., Whitlatch, R.B. & Osman, R.W. (2002). Linking climate change and biological invasions: Ocean warming facilitates nonindigenous species invasions. *Proc. Natl Acad. Sci. USA*, 99, 15497–15500.
- Stewart, G. (2010). Meta-analysis in applied ecology. Biol. Lett., 6, 78-81.
- Sturtz, S., Ligges, U. & Gelman, A. (2005). R2WinBUGS: A Package for Running WinBUGS from R. J. Stat. Softw., 12, 1–16.
- Sweeting, M.J., Sutton, A.J. & Lambert, P.C. (2004). What to add to nothing? Use and avoidance of continuity corrections in meta-analysis of sparse data. *Stat. Med.*, 23, 1351–1375.
- Terrestrial Carbon (TerraC) Information System. (2011). University of Florida, Gainesville. Available at: http://TerraC.ifas.ufl.edu. Last accessed 14 October 2011.
- Theoharides, K.A. & Dukes, J.S. (2007). Plant invasion across space and time: factors affecting nonindigenous species success during four stages of invasion. *New Phytol.*, 176, 256–273.
- Thomas, A., O'Hara, B., Ligges, U. & Sturtz, S. (2006). Making BUGS open. RNews, 6, 12–17. http://cran.r-project.org/doc/Rnews/>.

- Thuiller, W., Richardson, D.M., Pyšek, P., Midgley, G.F., Hughes, G.O. & Rouget, M. (2005). Niche-based modeling as a tool for predicting the risk of alien plant invasions at a global scale. *Glob. Change Biol.*, 11, 2234–2250.
- Thuiller, W., Richardson, D.M. & Midgley, G. (2007). Will climate change promote alien plant invasions? In *Biological Invasions*. (ed. Nentwig, W.). Springer-Verlag, Berlin, pp. 197–211.
- Vilà, M., Corbin, J.D., Dukes, J.S., Pino, J. & Smith, S.D. (2007). Linking plant invasions to global environmental change. In *Terrestrial Ecosystems in a Changing World.* (eds Canadell, J., Pataki, D. Pitelka, L.). Springer, New York, pp. 93–102.
- Walther, G.R., Post, E., Convey, P., Menzel, A., Parmesan, C., Beebee, T.J.C. et al. (2002). Ecological responses to recent climate change. Nature, 416, 389–395.
- Walther, G.R., Roques, A., Hulme, P.E., Sykes, M.T., Pyšek, P., Kuhn, I. *et al.* (2009). Alien species in a warmer world: risks and opportunities. *Trends Ecol. Evol.*, 24, 686–693.
- Webber, B.L. & Scott, J.K. (2012). Rapid global change: implications for defining natives and aliens. *Glob. Ecol. Biogeogr.*, 21, 305–311.
- Williams, S.L. & Smith, J.E. (2007). A global review of the distribution, taxonomy, and impacts of introduced seaweeds. *Ann. Rev. Ecol. Evol. Syst.*, 38, 327–359.
- Willis, C.G., Ruhfel, B.R., Primack, R.B., Miller-Rushing, A.J., Losos, J.B. & Davis, C.C. (2010). Favorable climate change response explains non-native species' success in Thoreau's woods. *PLoS ONE*, 5, e8878.
- Witte, S., Buschbaum, C., van Beusekom, J.E.E. & Reise, K. (2010). Does climatic warming explain why an introduced barnacle finally takes over after a lag of more than 50 years? *Biol. Invasions*, 12, 3579–3589.

- Wu, Z., Dijkstra, P., Koch, G.W., Peñuelas, J. & Hungate, B.A. (2011). Responses of terrestrial ecosystems to temperature and precipitation change: a meta-analysis of experimental manipulation. *Glob. Change Biol.*, 17, 927–942.
- Zerebecki, R.A. & Sorte, C.J.B. (2011). Temperature tolerance and stress proteins as mechanisms of invasive species success. *PLoS ONE*, 6, e14806.

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1 2 3	Appendix S1 Search terms for the systematic review and references for papers included in the meta- analysis.
3 4 5	Search terms
5 6 7 8	Each search included strings of descriptors for (1) non-native species, (2) taxa (in some cases by habitat), and (3) environmental drivers of climate change. Searches were carried out using ISI Web of Knowledge.
9 10	(1) Non-native species descriptors
11 12 13	invas* OR invad* OR alien OR non-nativ* OR nonnativ* OR exotic* OR introduced OR non-indigenous OR naturalized
14 15	(2) Taxonomic descriptors
16 17 18	Plants (terrestrial and aquatic): plant or vegetat* OR tree* OR shrub* OR grass* OR forest* OR forb* OR herb* OR vine*
19 20 21	Marine animals: marine OR ocean* OR sea OR intertidal OR subtidal OR mudflat OR estuarine OR pelagic OR saltmarsh
22 23	Freshwater animals: fish OR snail OR crayfish
24 25	Terrestrial animals: insect* OR beetle* OR mammal* OR bird* OR reptile* OR amphibian*
26 27	(3) Environmental driver descriptors
28 29	Temperature: temperature OR warm* OR heat*
30 31	CO ₂ : CO ₂ OR carbon dioxide
32 33	CO ₂ for aquatic studies: CO ₂ OR carbon dioxide OR pH OR ocean acidification
34 35	Precipitation: precip* OR rain* OR snow* OR water*
36 37	For example, a search for temperature impacts on terrestrial autotrophs was entered as:
38 39 40 41 42	TOPIC = (invas* OR invad* OR alien OR non-nativ* OR nonnativ* OR exotic* OR introduced OR non- indigenous OR naturalized OR naturalized) AND (temperature OR warm* OR heat*) AND (plant or vegetat* OR tree* OR shrub* OR grass* OR forest* OR forb* OR herb* OR vine*).
43 44	References
45 46 47 48 49	 Allen, E.B. (1982). Germination and Competition of <i>Salsola kali</i> with native C3 and C4 species under three temperature regimes. <i>Bull. Torrey Bot. Club</i>, 109, 39-46. Anderson, L.J., Maherali, H., Johnson, H.B., Polley, H.W. & Jackson, R.B. (2001). Gas exchange and photosynthetic acclimation over subambient to elevated CO₂ in a C3-C4 grassland. <i>Glob. Change Biol.</i>, 7, 693-707.
50 51	Arnone, J.A., Jasoni, R.L., Lucchesi, A.J., Larsen, J.D., Leger, E.A., Sherry, R.A., Luo, Y., Schimel, D.S. & Verburg, P.S.J. (2011). A climatically extreme year has large impacts on C4 species in tallgrass

- prairie ecosystems but only minor effects on species richness and other plant functional groups. *J. Ecol.*, 99, 678-688.
- Ashby, W.C. & Hellmers, H. (1959). Flowering and growth responses to photoperiod and temperature for
 six southern California grasses. *Bot. Gazette*, 120, 151-157.
- Bahrani, M.J., Bahrami, H. & Haghighi, A.A.K. (2010). Effect of water stress on ten forage grasses native
 or introduced to Iran. *Grassl. Sci.*, 56, 1-5.
- Bakker, J. & Wilson, S. (2001). Competitive abilities of introduced and native grasses. *Plant Ecol.*, 157, 119-127.
- Baruch, Z. & Jackson, R. (2005). Responses of tropical native and invader C₄ grasses to water stress,
 clipping and increased atmospheric CO₂ concentration. *Oecologia*, 145, 522-532.
- Belote, R.T., Weltzin, J.F. & Norby, R.J. (2004). Response of an understory plant community to elevated
 [CO₂] depends on differential responses of dominant invasive species and is mediated by soil
 water availability. *New Phytol.*, 161, 827-835.
- Blicker, P.S., Olson, B.E. & Wraith, J.M. (2003). Water use and water-use efficiency of the invasive
 Centaurea maculosa and three native grasses. *Plant Soil*, 254, 371-381.
- Blumenthal, D., Chimner, R.A., Welker, J.M. & Morgan, J.A. (2008). Increased snow facilitates plant
 invasion in mixedgrass prairie. *New Phytol.*, 179, 440-448.
- Boschma, S.P., Hill, M.J., Scott, J.M. & Rapp, G.G. (2003). The response to moisture and defoliation
 stresses, and traits for resilience of perennial grasses on the Northern Tablelands of New South
 Wales, Australia. *Anglais*, 54, 14.
- Braby, C.E. & Somero, G.N. (2006). Following the heart: temperature and salinity effects on heart rate in native and invasive species of blue mussels (genus *Mytilus*). J. Exper. Biol., 209, 2554-2566.
- Brock, M.T. & Galen, C. (2005). Drought tolerance in the alpine dandelion, *Taraxacum ceratophorum* (Asteraceaea), its exotic congener *T. officinale*, and interspecific hybrids under natural and
 experimental conditions. *Am. J. Bot.*, 92, 1311-1321.
- Buckland, S.M., Thompson, K., Hodgson, J.G. & Grime, J.P. (2001). Grassland invasions: effects of
 manipulations of climate and management. *J. Appl. Ecol.*, 38, 301-309.
- Bunting, D.P., Kurc, S.A. & Grabau, M.R. (2011). Using existing agricultural infrastructure for
 restoration practices: Factors influencing successful establishment of *Populus fremontii* over
 Tamarix ramosissima. J. Arid Environ., 75, 851-860.
- Cabin, R.J., Weller, S.G., Lorence, D.H., Cordell, S. & Hadway, L.J. (2002). Effects of microsite, water,
 weeding, and direct seeding on the regeneration of native and alien species within a Hawaiian dry
 forest preserve. *Biol. Conserv.*, 104, 181-190.
- Carveth, C.J., Widmer, A.M. & Bonar, S.A. (2006). Comparison of upper thermal tolerances of native
 and nonnative fish species in Arizona. *Trans. Am. Fish. Soc.*, 135, 1433-1440.
- Cox, J.A. & Conran, J.G. (1996). The effect of water stress on the life cycles of *Erodium crinitum* Carolin
 and *Erodium cicutarium* (L.) L'Hérit. ex Alton (Geraniaceae). *Aust. J. Ecol.*, 21, 235-240.
- Domènech, R. & Vilà, M. (2008). Response of the invader *Cortaderia selloana* and two coexisting
 natives to competition and water stress. *Biol. Invasions*, 10, 903-912.
- Dukes, J.S. (2002). Comparison of the effect of elevated CO₂ on an invasive species (*Centaurea* solstitialis) in monoculture and community settings. *Plant Ecol.*, 160, 225-234.
- Duyck, P.-F., David, P. & Quilici, S. (2006). Climatic niche partitioning following successive invasions
 by fruit flies in La Réunion. *J. Anim. Ecol.*, 75, 518-526.
- Engel, E.C., Weltzin, J.F., Norby, R.J. & Classen, A.T. (2009). Responses of an old-field plant
 community to interacting factors of elevated [CO₂], warming, and soil moisture. *J. Plant Ecol.*, 2,
 1-11.
- Findlay, H.S., Kendall, M.A., Spicer, J.I. & Widdicombe, S. (2010). Post-larval development of two
 intertidal barnacles at elevated CO₂ and temperature. *Mar. Biol.*, 157, 725-735.
- Funk, J. & Zachary, V. (2010). Physiological responses to short-term water and light stress in native and
 invasive plant species in southern California. *Biol. Invasions*, 12, 1685-1694.

- Garcia-Serrano, H., Josep, E. & Sans, F.X. (2004). Factors that limit the emergence and establishment of
 the related aliens *Senecio inaequidens* and *Senecio pterophorus* and the native *Senecio malacitanus* in Mediterranean climate. *Can. J. Bot.*, 82, 1346-1355.
- Geng, Y.-P., Pan, X.-Y., Xu, C.-Y., Zhang, W.-J., Li, B. & Chen, J.-K. (2006). Phenotypic plasticity
 of invasive *Alternanthera philoxeroides* in relation to different water availability, compared
 to its native congener. *Acta Oecol.*, 30, 380-385.
- 108 Gray, A.J. & Mogg, R.J. (2001). Climate impacts on pioneer saltmarsh plants. *Clim. Res.*, 18, 105-112.
- Gutiérrez, J.R. (1992). Effects of low water supplementation and nutrient addition on the aboveground
 biomass production of annual plants in a Chilean coastal desert site. *Oecologia*, 90, 556-559.
- Hättenschwiler, S. & Körner, C. (2003). Does elevated CO₂ facilitate naturalization of the non-indigenous
 Prunus laurocerasus in Swiss temperate forests? *Func. Ecol.*, 17, 778-785.
- Hellmers, H. & Ashby, W.C. (1958). Growth of native and exotic plants under controlled temperatures
 and in the San Gabriel mountains California. *Ecology*, 39, 416-428.
- Hely, S.E.L. & Roxburgh, S.H. (2005). The interactive effects of elevated CO₂ temperature and initial
 size on growth and competition between a native C₃ and an invasive C₃ grass. *Plant Ecol.*, 177,
 85-98.
- Hill, J.P., Germino, M.J., Wraith, J.M., Olson, B.E. & Swan, M.B. (2006). Advantages in water relations
 contribute to greater photosynthesis in *Centaurea maculosa* compared with established grasses.
 Int. J. Plant Sci., 167, 269-277.
- Holway, D.A., Suarez, A.V. & Case, T.J. (2002). Role of abiotic factors in governing susceptibility to
 invasion: a test with Argentine ants. *Ecology*, 83, 1610-1619.
- Horton, J.L. & Clark, J.L. (2001). Water table decline alters growth and survival of *Salix gooddingii* and
 Tamarix chinensis seedlings. *For. Ecol. Manag.*, 140, 239-247.
- Hoya, A., Shibaike, H., Morita, T. & Ito, M. (2004). Germination and seedling survivorship
 characteristics of hybrids between native and alien species of dandelion (*Taraxacum*). *Plant Species Biol.*, 19, 81-90.
- Huxman, T. & Smith, S. (2001). Photosynthesis in an invasive grass and native forb at elevated CO₂
 during an El Niño year in the Mojave Desert. *Oecologia*, 128, 193-201.
- Hwang, B. & Lauenroth, W. (2008). Effect of nitrogen, water and neighbor density on the growth of
 Hesperis matronalis and two native perennials. *Biol. Invasions*, 10, 771-779.
- Ignace, D.D., Juxman, T.E., Weltzin, J.F. & Williams, D.G. (2007). Leaf gas exchange and water status
 responses of a native and non-native grass to precipitation across contrasting soil surfaces in the
 Sonoran desert. *Oecologia*, 152, 401-413.
- Janion, C., Leinaas, H.P., Terblanche, J.S. & Chown, S.L. (2010). Trait means and reaction norms: the
 consequences of climate change/invasion interactions at the organism level. *Evol. Ecol.*, 24,
 1365-1380.
- Kim, H.-R. & You, Y.-H. (2010). Effects of elevated CO₂ concentration and increased temperature on
 leaf related-physiological responses of *Phytolacca insularis* (native species) and *Phytolacca americana* (invasive species). *J. Ecol. Field Biol.*, 33, 195-204.
- Leishman, M.R. & Thomson, V.P. (2005). Experimental evidence for the effects of additional water,
 nutrients and physical disturbance on invasive plants in low fertility Hawkesbury Sandstone soils,
 Sydney, Australia. J. Ecol., 93, 38-49.
- Lenssen, G.M., Lamers, J., Stroetenga, M. & Rozema, J. (1993). Interactive effects of atmospheric CO₂
 enrichment, salinity and flooding on growth of C₃ (*Elymus athericus*) and C₄ (*Spartina anglica*)
 salt marsh species. *Plant Ecol.*, 104-105, 379-388.
- Levine, J.M., McEachern, A.K. & Cowan, C. (2010). Do competitors modulate rare plant response to
 precipitation change? *Ecology*, 91, 130-140.
- Manea, A. & Leishman, M. (2011). Competitive interactions between native and invasive exotic plant
 species are altered under elevated carbon dioxide. *Oecologia*, 165, 735-744.
- Mangla, S., Sheley, R.L. & James, J.J. (2011). Field growth comparisons of invasive alien annual and
 native perennial grasses in monocultures. J. Arid Environ., 75, 206-210.

- Maron, John L. & Marler, M. (2008). Effects of native species diversity and resource additions on invader
 impact. Am. Nat., 172, S18-S33.
- Miller, A.W., Reynolds, A.C., Sobrino, C. & Riedel, G.F. (2009). Shellfish face uncertain future in high
 CO₂ world: Influence of acidification on oyster larvae calcification and growth in estuaries. *PLoS ONE*, 4, e5661.
- Morecroft, M.D., Stokes, V.J., Taylor, M.E. & Morison, J.I.L. (2008). Effects of climate and management
 history on the distribution and growth of sycamore (*Acer pseudoplatanus* L.) in a southern British
 woodland in comparison to native competitors. *Forestry*, 81, 59-74.
- Nagel, J.M., Huxman, T.E., Griffin, K.L. & Smith, S.D. (2004). CO₂ enrichment reduces the energetic
 cost of biomass construction in an invasive desert grass. *Ecology*, 85, 100-106.
- Naumann, J., Bissett, S., Young, D., Edwards, J. & Anderson, J. (2010). Diurnal patterns of
 photosynthesis, chlorophyll fluorescence, and PRI to evaluate water stress in the invasive species,
 Elaeagnus umbellata Thunb. *Trees Struct. Func.*, 24, 237-245.
- Nernberg, D. & Dale, M.R.T. (1997). Competition of five native prairie grasses with *Bromus inermis* under three moisture regimes. *Can. J. Bot.*, 75, 2140-2145.
- Parker, L.M., Ross, P.M. & O'Connor, W.A. (2010). Comparing the effect of elevated pCO₂ and
 temperature on the fertilization and early development of two species of oysters. *Mar. Biol.*, 157, 2435-2452.
- Patterson, D.T. (1993). Effects of day and night temperature on goatsrue (*Galega officinalis*) and alfalfa
 (*Medicago sativa*) growth. *Weed Sci.*, 41, 38-45.
- Patterson, D.T., Russell, A.E., Mortensen, D.A., Coffin, R.D. & Flint, E.P. (1986). Effects of temperature
 and photoperiod on Texas panicum (*Panicum texanum*) and wild proso millet (*Panicum miliaceum*). Weed Sci., 34, 876-882.
- Petes, L.E., Menge, B.A. & Murphy, G.D. (2007). Environmental stress decreases survival, growth, and
 reproduction in New Zealand mussels. *J. Exper. Mar. Biol. Ecol.*, 351, 83-91.
- Raizada, P., Singh, A. & Raghubanshi, A.S. (2009). Comparative response of seedlings of selected native
 dry tropical and alien invasive species to CO₂ enrichment. *J. Plant Ecol.*, 2, 69-75.
- Rao, L. & Allen, E. (2010). Combined effects of precipitation and nitrogen deposition on native and
 invasive winter annual production in California deserts. *Oecologia*, 162, 1035-1046.
- Reich, P.B., Tilman, D., Craine, J., Ellsworth, D., Tjoelker, M.G., Knops, J., Wedin, D., Naeem, S.,
 Bahauddin, D., Goth, J., Bengtson, W. & Lee, T.D. (2001). Do species and functional groups
 differ in acquisition and use of C, N and water under varying atmospheric CO₂ and N availability
 regimes? A field test with 16 grassland species. *New Phytol.*, 150, 435-448.
- Reynolds, L.V. & Cooper, D.J. (2010). Environmental tolerance of an invasive riparian tree and its
 potential for continued spread in the southwestern US. J. Veg. Sci., 21, 733-743.
- Sareyka, J., Kraufvelin, P., Lenz, M., Lindström, M., Tollrian, R. & Wahl, M. (2011). Differences in
 stress tolerance and brood size between a non-indigenous and an indigenous gammarid in the
 northern Baltic Sea. *Mar. Biol.*, 158, 2001-2008.
- Sasek, T.W. & Strain, B.R. (1991). Effects of CO₂ enrichment on the growth and morphology of a native
 and an introduced honeysuckle vine. *Am. J. Bot.*, 78, 69-75.
- Schneider, K.R. (2008). Heat stress in the intertidal: comparing survival and growth of an invasive and
 native mussel under a variety of thermal conditions. *Biol. Bull.*, 215, 253-264.
- Schneider, K.R. & Helmuth, B.S.T. (2007). Spatial variability in habitat temperature may drive patterns
 of selection between an invasive and native mussel species. *Mar. Ecol. Prog. Ser.*, 339, 157-167.
- Schumacher, E., Kueffer, C., Tobler, M., Gmür, V., Edwards, P.J. & Dietz, H. (2008). Influence of
 drought and shade on seedling growth of native and invasive trees in the Seychelles. *Biotropica*,
 40, 543-549.
- Smith, S.D., Huxman, T.E., Zitzer, S.F., Charlet, T.N., Housman, D.C., Coleman, J.S., Fenstermaker,
 L.K., Seemann, J.R. & Nowak, R.S. (2000). Elevated CO₂ increases productivity and invasive
 species success in an arid ecosystem. *Nature*, 408, 79-82.

- Song, L., Wu, J., Li, C., Li, F., Peng, S. & Chen, B. (2009). Different responses of invasive and native
 species to elevated CO₂ concentration. *Acta Oecol.*, 35, 128-135.
- Song, L.-Y., Li, C.-H. & Peng, S.-L. (2010). Elevated CO₂ increases energy-use efficiency of invasive
 Wedelia trilobata over its indigenous congener. *Biol. Invasions*, 12, 1221-1230.
- Sorte, C.J.B., Fuller, A. & Bracken, M.E.S. (2010). Impacts of a simulated heat wave on composition of a
 marine community. *Oikos*, 119, 1909-1918.
- Sorte, C.J.B., Williams, S.L. & Zerebecki, R.A. (2010). Ocean warming increases threat of invasive
 species in a marine fouling community. *Ecology*, 91, 2198-2204.
- Steen, H. & Rueness, J. (2004). Comparison of survival and growth in germlings of six fucoid species
 (Fucales, Phaeophyceae) at two different temperature and nutrient levels. *Sarsia*, 89, 175-183.
- Steiner, J.J., Brewer, T.G. & Griffith, S.M. (2001). Temperature effects on interspecific interference
 among two native wetland grasses and tall fescue. *Agron. J.*, 93, 1020-1027.
- Sullivan, L., Wildova, R., Goldberg, D. & Vogel, C. (2010). Growth of three cattail (*Typha*) taxa in
 response to elevated CO₂. *Plant Ecol.*, 207, 121-129.
- Sun, Y., Ding, J. & Frye, M.J. (2010). Effects of resource availability on tolerance of herbivory in the
 invasive Alternanthera philoxeroides and the native Alternanthera sessilis. Weed Res., 50, 527 536.
- Tang, Y.Q. & Yokomi, R.K. (1995). Temperature-dependent development of three hymenopterous
 parasitoids of aphids (Homoptera: Aphididae) attacking citrus. *Environ. Entomol.*, 24, 1736-1740.
- Verlinden, M. & Nijs, I. (2010). Alien plant species favoured over congeneric natives under experimental
 climate warming in temperate Belgian climate. *Biol. Invasions*, 12, 2777-2787.
- Werner, C., Zumkier, U., Beyschlag, W. & Máguas, C. (2010). High competitiveness of a resource
 demanding invasive acacia under low resource supply. *Plant Ecol.*, 206, 83-96.
- Williams, A.L., Wills, K.E., Janes, J.K., Vander Schoor, J.K., Newton, P.C.D. & Hovenden, M.J. (2007).
 Warming and free-air CO₂ enrichment alter demographics in four co-occurring grassland species.
 New Phytol., 176, 365-374.
- Williams, D.G. & Black, R.A. (1994). Drought response of a native and introduced Hawaiian grass.
 Oecologia, 97, 512-519.
- Willis, A.J., Berentson, P.R. & Ash, J.E. (2003). Impacts of a weed biocontrol agent on recovery from
 water stress in a target and a non-target *Hypericum* species. *J. Appl. Ecol.*, 40, 320-333.
- Wilson, S.B., Wilson, P.C. & Albano, J.A. (2004). Growth and development of the native *Ruellia caroliniensis* and invasive *Ruellia tweediana*. *HortScience*, 39, 1015-1019.
- Wu, Y.P., Hu, X.W. & Wang, Y.R. (2009). Growth, water relations, and stomatal development of
 Caragana korshinskii Kom. and *Zygophyllum xanthoxylum* (Bunge) Maxim. seedlings in
 response to water deficits. *New Zealand J. Agric. Res.*, 52, 185-193.
- Xu, B., Li, F., Shan, L.U.N., Ma, Y., Ichizen, N. & Huang, J.I.N. (2006). Gas exchange, biomass
 partition, and water relationships of three grass seedlings under water stress. *Weed Biol. Manag.*,
 6, 79-88.
- Yazaki, K., Sano, Y., Fujikawa, S., Nakano, T. & Ishida, A. (2010). Response to dehydration and
 irrigation in invasive and native saplings: osmotic adjustment versus leaf shedding. *Tree Physiol.*,
 30, 597-607.
- Zerebecki, R.A. & Sorte, C.J.B. (2011). Temperature tolerance and stress proteins as mechanisms of
 invasive species success. *PLoS ONE*, 6, e14806.
- Zhang, C., Wei, L., ZhiFang, X., HongLin, C. & WanHui, Y. (2006). Responses of vegetative growth and
 photosynthesis to temperature in the invasive species *Alternanthera philoxeroides* and its
 indigenous congener *A. sessilis. J. Trop. Subtrop. Bot.*, 14, 333-339.
- Zukowski, S. & Walker, K.F. (2009). Freshwater snails in competition: alien *Physa acuta* (Physidae) and
 native *Glyptophysa gibbosa* (Planorbidae) in the River Murray, South Australia. *Mar. Freshw. Res.*, 60, 999-1005.

Appendix S2 Table summarizing studies included in the meta-analysis. Note that individual papers could include multiple experiments (e.g., Exp. a, b, etc.), and the sample size for studies that did not report individual (but rather composite) species responses is designated as a "group". For references, see App. S1.

Increased temperature

Reference	Exp.	System	Taxon	Habitat	Study location	Response type	N _{non-native}	N _{native}
Braby & Somero 2006	a	Aquatic	Invertebrate	Subtidal	California, USA	Survival	1	1
Braby & Somero 2006	b	Aquatic	Invertebrate	Subtidal	California, USA	Survival	1	1
Braby & Somero 2006	c	Aquatic	Invertebrate	Subtidal	California, USA	Survival	1	1
Carveth et al. 2006	а	Aquatic	Fish	Stream	Arizona, USA	Survival	4	10
Findlay et al. 2010	а	Aquatic	Invertebrate	Intertidal	England	Growth, Survival	1	1
Parker et al. 2010	а	Aquatic	Invertebrate	Subtidal	Australia	Growth	1	1
Parker et al. 2010	b	Aquatic	Invertebrate	Subtidal	Australia	Growth	1	1
Parker et al. 2010	с	Aquatic	Invertebrate	Subtidal	Australia	Survival	1	1
Petes et al. 2007	а	Aquatic	Invertebrate	Intertidal	New Zealand	Growth	1	1
Sareyka et al. 2011	а	Aquatic	Invertebrate	Subtidal	Finland	Survival	1	1
Sareyka et al. 2011	b	Aquatic	Invertebrate	Subtidal	Finland	Survival	1	1
Schneider & Helmuth 2007	a	Aquatic	Invertebrate	Intertidal	California, USA	Survival	1	1
Schneider 2008	а	Aquatic	Invertebrate	Subtidal	Washington, USA	Growth	1	1
Schneider 2008	b	Aquatic	Invertebrate	Subtidal	Washington, USA	Growth	1	1
Sorte et al. 2010a	а	Aquatic	Invertebrate	Subtidal	California, USA	Survival	6	2
Sorte et al. 2010b	а	Aquatic	Invertebrate	Subtidal	California, USA	Growth	5	1
Sorte et al. 2010b	b	Aquatic	Invertebrate	Subtidal	California, USA	Survival	6	1
Steen & Rueness 2004	а	Aquatic	Seaweed	Subtidal	Norway	Survival	1	5
Zerebecki & Sorte 2011	а	Aquatic	Invertebrate	Subtidal	California, USA	Survival	6	2
Zukowsji & Walker 2009	а	Aquatic	Invertebrate	Stream	Australia	Growth	1	1
Allen 1982	а	Terrestrial	Plant	Grassland	Wyoming, USA	Growth	1	4
Arnone et al. 2011	a	Terrestrial	Plant	Grassland	Oklahoma / Nevada, USA	Growth	2	14
Ashby & Hellmers 1959	а	Terrestrial	Plant	Shrubland	California, USA	Growth	2	4
Blumenthal & Perry <i>unpubl. data</i>	b	Terrestrial	Plant	Grassland	Wyoming, USA	Growth, Photosyn.	3	4
Blumenthal & Perry <i>unpubl. data</i>	c	Terrestrial	Plant	Grassland	Wyoming, USA	Growth, Photosyn.	3	4

Buckland et al. 2001	а	Terrestrial	Plant	Grassland	England	Growth	1	1
Duyck et al. 2006	а	Terrestrial	Insect	n/a	La Reunion, Indian Ocean	Survival	3	1
Engel et al. 2009	а	Terrestrial	Plant	Herbaceous	Tennessee, USA	Growth	5	2
Gray & Mogg 2001	а	Terrestrial	Plant	Wetland	England	Growth	1	1
Hellmers & Ashby 1958	а	Terrestrial	Plant	Forest	California, USA	Growth	21	6
Hely & Roxburgh 2005	а	Terrestrial	Plant	Grassland	Australia	Growth	1	1
Hely & Roxburgh 2005	b	Terrestrial	Plant	Grassland	Australia	Growth	1	1
Holway et al. 2002	b	Terrestrial	Insect	Shrubland	California, USA	Survival	1	5
Hoya <i>et al</i> . 2004	а	Terrestrial	Plant	Herbaceous	Japan	Growth	1	1
Janion et al. 2010	а	Terrestrial	Insect	Tundra	Marion Is., Southern Ocean	Reproduction	4	2
Kim & You 2010	а	Terrestrial	Plant	Herbaceous	Korea	Photosynthesis	1	1
Naumann et al. 2010	а	Terrestrial	Plant	Forest	Virginia, USA	Photosynthesis	1	1
Patterson 1993	а	Terrestrial	Plant	Grassland	Utah, USA	Growth	1	1
Patterson et al. 1986	a	Terrestrial	Plant	Grassland	Minnesota / Georgia, USA	Growth	1	1
Steiner et al. 2001	а	Terrestrial	Plant	Grassland	Oregon, USA	Growth	1	2
Tang & Yokomi 1995	а	Terrestrial	Insect	Forest	Florida, USA	Growth	2	1
Verlinden & Nijs 2010	а	Terrestrial	Plant	Herbaceous	Belgium	Growth	10	10
Williams et al. 2007	b	Terrestrial	Plant	Grassland	Tasmania, Australia	Growth	2	2
Williams et al. 2007	c	Terrestrial	Plant	Grassland	Tasmania, Australia	Growth	2	2
Wilson et al. 2004	а	Terrestrial	Plant	Herbaceous	Florida, USA	Growth	1	1
Zhang et al. 2006	а	Terrestrial	Plant	Herbaceous	China	Growth	1	1

Increased CO₂

Reference	Exp.	System	Taxon	Habitat	Study location	Response type	$N_{ m non-native}$	N _{native}
Findlay et al. 2010	b	Aquatic	Invertebrate	Intertidal	England	Growth, Survival	1	1
Lenssen et al. 1993	а	Aquatic	Plant	Herbaceous	The Netherlands	Growth	1	1
Lenssen et al. 1993	b	Aquatic	Plant	Herbaceous	The Netherlands	Growth	1	1
Miller et al. 2009	а	Aquatic	Invertebrate	Wetland	Maryland, USA	Growth	1	1
Parker et al. 2010	а	Aquatic	Invertebrate	Subtidal	Australia	Growth	1	1
Parker et al. 2010	b	Aquatic	Invertebrate	Subtidal	Australia	Growth	1	1
Parker et al. 2010	c	Aquatic	Invertebrate	Subtidal	Australia	Survival	1	1
Sullivan et al. 2010	а	Aquatic	Plant	Wetland	Michigan, USA	Growth	1	1

Anderson et al. 2001	а	Terrestrial	Plant	Herbaceous	Texas, USA	Growth	2	1
Baruch & Jackson 2005	а	Terrestrial	Plant	Grassland	Venezuela	Growth	2	1
Baruch & Jackson 2005	b	Terrestrial	Plant	Grassland	Venezuela	Growth	2	1
Belote et al. 2004	а	Terrestrial	Plant	Herbaceous	Tennessee, USA	Growth	2	3
Blumenthal & Perry <i>unpubl. data</i>	а	Terrestrial	Plant	Grassland	Wyoming, USA	Growth, Photosyn.	3	4
Blumenthal & Perry <i>unpubl. data</i>	c	Terrestrial	Plant	Grassland	Wyoming, USA	Growth, Photosyn.	3	4
Dukes 2002	а	Terrestrial	Plant	Herbaceous	California, USA	Growth	1	1
Hättenschwiler & Körner 2003	а	Terrestrial	Plant	Forest	Switzerland	Growth	1	4
Huxman & Smith 2001	а	Terrestrial	Plant	Desert	Nevada, USA	Growth	1	1
Kim & You 2010	а	Terrestrial	Plant	Herbaceous	Korea	Photosynthesis	1	1
Manea & Leishman 2011	а	Terrestrial	Plant	Forest	Australia	Growth	14	14
Manea & Leishman 2011	b	Terrestrial	Plant	Forest	Australia	Growth	14	14
Nagel et al. 2004	а	Terrestrial	Plant	Desert	Nevada, USA	Growth	1	2
Naumann et al. 2010	а	Terrestrial	Plant	Forest	Virginia, USA	Photosynthesis	1	1
Perry unpubl. data	а	Terrestrial	Plant	Forest	Colorado, USA	Growth	3	2
Raizada et al. 2009	а	Terrestrial	Plant	Forest	India	Growth	2	4
Reich et al. 2001	а	Terrestrial	Plant	Grassland	Minnesota, USA	Growth	4	12
Sasek & Strain 1991	а	Terrestrial	Plant	Herbaceous	North Carolina, USA	Growth	1	1
Smith <i>et al.</i> 2000	а	Terrestrial	Plant	Desert	Nevada, USA	Reproduction	1	2
Song <i>et al.</i> 2009	а	Terrestrial	Plant	Forest	China	Growth	3	3
Song <i>et al.</i> 2010	а	Terrestrial	Plant	Forest	China	Growth	1	1
Williams et al. 2007	а	Terrestrial	Plant	Grassland	Tasmania, Australia	Growth	2	2
Williams et al. 2007	c	Terrestrial	Plant	Grassland	Tasmania, Australia	Growth	2	2

Increased Precipitation

Reference	Exp.	System	Taxon	Habitat	Study location	Response type	$N_{ m non-native}$	N _{native}
Bakker & Wilson 2001	b	Terrestrial	Plant	Herbaceous	Saskatchewan, Canada	Growth	1	1
Bakker & Wilson 2001	d	Terrestrial	Plant	Herbaceous	Saskatchewan, Canada	Growth	1	1
Blicker et al. 2003	b	Terrestrial	Plant	Grassland	Montana, USA	Growth	1	3

Blumenthal et al. 2008	а	Terrestrial	Plant	Grassland	Wyoming, USA	Growth	3	group
Brock & Galen 2005	а	Terrestrial	Plant	Tundra	Colorado, USA	Photosynthesis	1	1
Cabin <i>et al.</i> 2002	а	Terrestrial	Plant	Forest	Hawaii, USA	Growth	3	7
Coffman unpubl. data	а	Terrestrial	Plant	Forest	California, USA	Growth	1	3
Garcia-Serrano <i>et al.</i> 2004	а	Terrestrial	Plant	Grassland	Spain	Survival	2	1
Garcia-Serrano <i>et al.</i> 2004	b	Terrestrial	Plant	Grassland	Spain	Survival	2	1
Garcia-Serrano <i>et al.</i> 2004	с	Terrestrial	Plant	Forest	Spain	Survival	2	1
Geng et al. 2006	b	Terrestrial	Plant	Herbaceous	China	Growth	1	1
Gutiérrez 1992	а	Terrestrial	Plant	Grassland	Chile	Growth	4	3
Gutiérrez 1992	b	Terrestrial	Plant	Grassland	Chile	Growth	2	3
Hill et al. 2006	b	Terrestrial	Plant	Grassland	Montana, USA	Photosynthesis	1	2
Holway et al. 2002	а	Terrestrial	Insect	Shrubland	California, USA	Survival	1	1
Leishman & Thomson 2005	а	Terrestrial	Plant	Shrubland	Australia	Growth, Survival	group	group
Levine et al. 2010	b	Terrestrial	Plant	Grassland	California, USA	Growth	1	1
Maron & Marler 2008	а	Terrestrial	Plant	Herbaceous	Montana, USA	Growth	3	10
Naumann et al. 2010	а	Terrestrial	Plant	Forest	Virginia, USA	Photosynthesis	1	1
Nernberg & Dale 1997	b	Terrestrial	Plant	Grassland	Saskatchewan, Canada	Growth	1	5
Rao & Allen 2010	b	Terrestrial	Plant	Herbaceous	California, USA	Growth	1	1
Reynolds & Cooper 2010	b	Terrestrial	Plant	Forest	Colorado, USA	Growth, Survival	2	1
Reynolds & Cooper 2010	d	Terrestrial	Plant	Forest	Colorado, USA	Growth, Survival	2	1

Decreased Precipitation

Reference	Exp.	System	Taxon	Habitat	Study location	Response type	N _{non-native}	N _{native}
Bahrani et al. 2010	a	Terrestrial	Plant	Grassland	Iran	Growth	3	7
Bakker & Wilson 2001	a	Terrestrial	Plant	Herbaceous	Saskatchewan, Canada	Growth	1	1
Bakker & Wilson 2001	c	Terrestrial	Plant	Herbaceous	Saskatchewan, Canada	Growth	1	1
Baruch & Jackson 2005	a	Terrestrial	Plant	Grassland	Venezuela	Growth	2	1
Baruch & Jackson 2005	b	Terrestrial	Plant	Grassland	Venezuela	Growth	2	1

Blicker et al. 2003	а	Terrestrial	Plant	Grassland	Montana, USA	Growth	1	3
Boschma et al. 2003	а	Terrestrial	Plant	Herbaceous	Australia	Growth	4	2
Buckland et al. 2001	b	Terrestrial	Plant	Grassland	England	Growth	1	1
Bunting et al. 2011	а	Terrestrial	Plant	Forest	Arizona, USA	Growth, Survival	1	1
Bunting et al. 2011	b	Terrestrial	Plant	Forest	Arizona, USA	Growth, Survival	1	1
Cox & Conran 1996	а	Terrestrial	Plant	Shrubland	Australia	Reproduction	1	1
Domenech & Vilà 2008	а	Terrestrial	Plant	Grassland	Spain	Growth	1	2
Duyck et al. 2006	b	Terrestrial	Insect	n/a	La Reunion, Indian Ocean	Survival	3	1
Funk & Zachary 2010	а	Terrestrial	Plant	Shrubland	California, USA	Photosynthesis	2	2
Geng et al. 2006	а	Terrestrial	Plant	Herbaceous	China	Growth	1	1
Hill et al. 2006	а	Terrestrial	Plant	Grassland	Montana, USA	Photosynthesis	1	2
Horton & Clark 2001	а	Terrestrial	Plant	Forest	Arizona, USA	Survival	1	1
Hwang & Lauenroth 2008	а	Terrestrial	Plant	Herbaceous	Colorado, USA	Growth	1	2
Ignace et al. 2007	а	Terrestrial	Plant	Herbaceous	Arizona, USA	Growth	1	1
Ignace et al. 2007	b	Terrestrial	Plant	Herbaceous	Arizona, USA	Growth	1	1
Levine et al. 2010	а	Terrestrial	Plant	Grassland	California, USA	Growth	1	1
Mangla et al. 2010	а	Terrestrial	Plant	Shrubland	Oregon, USA	Growth	2	1
Morecroft et al. 2008	а	Terrestrial	Plant	Forest	England	Growth	1	2
Nernberg & Dale 1997	а	Terrestrial	Plant	Grassland	Saskatchewan, Canada	Growth	1	5
Rao & Allen 2010	а	Terrestrial	Plant	Herbaceous	California, USA	Growth	1	1
Reynolds & Cooper 2010	а	Terrestrial	Plant	Forest	Colorado, USA	Growth	2	1
Reynolds & Cooper 2010	с	Terrestrial	Plant	Forest	Colorado, USA	Growth, Survival	2	1
Schumacher et al. 2008	a	Terrestrial	Plant	Forest	Republic of Seychelles	Growth	group	group
Sun et al. 2010	а	Terrestrial	Plant	Herbaceous	China	Growth	1	1
Werner et al. 2010	а	Terrestrial	Plant	Shrubland	Portugal	Growth	1	2
Williams & Black 1994	а	Terrestrial	Plant	Grassland	Hawaii, USA	Growth	1	1
Willis et al. 2003	а	Terrestrial	Plant	Grassland	Australia	Growth, Reprod.	1	1
Wu et al. 2009	а	Terrestrial	Plant	Shrubland	China	Growth	1	1
Xu et al. 2006	а	Terrestrial	Plant	Herbaceous	China	Growth	1	1
Yazaki <i>et al.</i> 2010	а	Terrestrial	Plant	Forest	Japan	Photosynthesis	1	1

Appendix S3 Results of variance-weighted meta-analysis.

To parallel the bootstrapped study-level analysis, we carried out a parameteric, variance-weighted metaanalysis at the species level using the difference-to-mean ratio effect size *ES* for the subset of studies that provided appropriate standard deviation and sample size information.



Figure S3.1 Results of variance-weighted analysis including the subset of studies for which variance data were available. Values shown are for the mildest treatment level from the 16 studies that included multiple treatment levels (note: results did not differ – visibly or statistically – when using the most extreme climate treatment for the studies that included 3+ levels). Error bars represent 95% confidence limits, and asterisks denote groups where *ES* was significantly different from zero. Individual species and separate studies within the same paper were treated as independent samples (Gurevitch & Hedges 2001). Comparisons to the control treatment for each study were determined using a mixed effects model with origin (native vs. non-native) treated as a fixed effect, using the metafor package (Viechtbauer 2010) in R 2.14 (R Development Core Team 2011). The number of species and studies used for each comparison is noted along the bottom of the figure.

References:

- Gurevitch, J. & Hedges, L.V. (2001). Meta-analysis Combining the results of independent experiments. Pp. 347-369 In: *Design and Analysis of Ecological Experiments*. Eds: Scheiner, S.M. & Gurevitch, J. Oxford: Oxford University Press.
- R Development Core Team (2011). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at: http://www.R-project.org/.
- Viechtbauer, W. (2010). Conducting meta-analyses in R with the metafor package. J. Stat. Software, 36, 1-48.



Appendix S4 Methods for the hierarchical analysis of response as a function of magnitude of the climate treatment.

Figure S4.1 Graphical representation of the hierarchical model's (a) structure and (b) functional relationship of analysis of effect size as a function of magnitude of treatment. θ . vector of parameters included in the analysis (a: maximum effect size and b: half-saturation constant), o: origin (native and non-native), t: terrestrial, a: aquatic, temp: temperature, prec: precipitation.

Supplementary methods for the hierarchical analysis

We explored the relationship between effect size (i.e., magnitude of performance response) and the treatment magnitude (i.e., degree of climatic change) within a hierarchical framework. For species *i* in study s (analyzed separately for negative and positive responses), the likelihood of observing that effect size was calculated as:

$$ES_{obs\ i,s} \sim Normal(ES_{i,s}, \sigma_i^2)$$

with process model:

 MT_s

 $ES_{i,s} = a_{origin (i),system (i),driver (i)} \overline{MT_s + b_{origin (i),system (i),driver (i)}}$

The variance associated with each effect size, σ_i^2 , was estimated as a combination of the observed variance in response size and an overall variance: $\frac{1}{\sigma_i^2} = \frac{1}{\sigma_{abs(i)}^2} + \frac{1}{\eta^2} \cdot \sigma_{abs(i)}^2$, the observed variance, was

calculated from the standard deviation (SD) or standard error reported in the original study for the response variable. To estimate the observed variance associated with each effect size, we ran 10,000 simulations calculating effect size from the mean response and reported SD. If measurements of variability around each effect size were not available (219 out of 755 observations), we then estimated this variance as $1/\sigma_{obs(i)}^2 \sim Gamma(0.01, 0.01)$. The overall variance, η^2 , was estimated from a distribution with non-informative priors, $1/\eta^2 \sim Gamma(0.01, 0.01)$.

The parameters, associated with the process model, for maximum effect size (parameter a) and the half saturation constant (parameter b) were estimated hierarchically as:

$$a_{origin,system,driver} \sim Normal(a1_{origin,system},\sigma_a^2)$$

 $a1_{origin,system} \sim Normal(a2_{origin},\sigma_{a1}^2)$ and $\sigma_a \sim Uniform(0,1000)$
 $a2_{origin} \sim LogNormal(0,10000)$ and $\sigma_{a1} \sim Uniform(0,1000)$

and

$$b_{origin,system,driver} \sim Normal(b1_{origin,system},\sigma_b^2)$$

 $b1_{origin,system} \sim Normal(b2_{origin}, \sigma_{b1}^2)$ and $\sigma_b \sim Uniform(0,1000)$
 $b2_{origin} \sim LogNormal(0,10000)$ and $\sigma_{b1} \sim Uniform(0,1000)$

We used non-informative priors for the hyperparameters, a^2 and b^2 , and the variances, σ^2 , and lognormal distributions for the overall parameters, a^2 and b^2 , to ensure positive values. This structure allowed us to make comparisons between native and non-native species at three levels: overall, parameters a^2_{origin} and

 $b2_{origin}$; within each system, parameters $a2_{origin,system}$ and $b2_{origin,system}$; and within system for each driver, parameters $a2_{origin,system,driver}$ and $b2_{origin,system,driver}$. Additional fixed (e.g., response type, latitude, or study duration) or random effects (e.g., study) did not improve the model fit (based on Deviance Information Criterion; Spiegelhalter *et al.* 2000); thus, results from the basic model were reported in the main text.

Reference

Spiegelhalter, D.J., Best, N., Carlin, B.P. & Linde, A.V.D. (2000). Bayesian measures of model complexity and fit. *J. Royal Statist. Soc. B*, 64, 583-639.

Appendix S5 Results of mixed models used to assess effects of potential covariates.

All mixed models were run in OpenBUGS, called from R using the R2OpenBUGS package. Three chains were run for 10,000 iterations, thinning by 25, and convergence was assessed visually and using the Gelman-Rubin criteria.

Model 1: The first mixed model was the simplest, modeling effect size (difference-to-mean ratio *ES*) as a function of origin-driver-system groups and including a random effect to account for studies.

ES ~ alpha_[Origin, driver, System] + RandomEffect_[study]



Figure S5.1: Results for Model 1. Points represent mean responses (difference-to-mean ratio *ES*; see *Methods*) for each origin-driver-system combination, and error bars represent 95% credible intervals. Asterisks denote responses that are significantly different from zero for native (black circles) and non-native species (gray triangles). The number under each pair of responses gives the probability that the non-native species are responding more positively than the natives (calculated from the difference between the two in the model). Clover symbols mark those where non-natives show a statistically significant advantage over native species.

Model 2: The second mixed model tested how effect size (difference-to-mean ratio *ES*) varied as a function of origin-driver-system groups and the magnitude of study treatment, while again including a random effect to account for studies.

ES ~ alpha_[Origin, driver, System] + beta_[Origin, driver, System]* Treatment Magnitude + RandomEffect_[study]



Figure S5.2. Intercepts (a) and coefficients (b) describing the effect of study magnitude on responses in each of the driver-origin categories. Points represent mean parameter estimates for each origin-driver-system combination, and intervals are 95% credible intervals. Asterisks denote effect sizes that are significantly different from zero for native (black circles) and non-native species (gray triangles). The number under each pair of responses gives the probability that the non-natives are responding more positively than the native species (calculated from the difference between the two in the model). Clover symbols mark comparisons where non-native species show a statistically significant advantage over natives.

Model 3. The third mixed model explored the effects of multiple additional covariates, none of which were significant in this analysis:

Random effects:

- Latitude
- Study duration
- Study magnitude
- Study

Fixed effects:

- Stage (Adult / Juvenile / Other)
- Habitat (Forest / Grassland / Herbaceous / Aquatic / Other)
- Response variable (Growth / Photosynth / Reproduction / Survival)
- Life history (Annual / Perennial / NA)



Figure S5.3. Parameter estimates for additional covariates. Points represent mean parameter estimates for each origin-driver-system combination, and intervals are 95% credible intervals.



Figure S5.3 (cont.). Parameter estimates for additional covariates. Points represent mean parameter estimates for each origin-driver-system combination, and intervals are 95% credible intervals.



Figure S5.4. Contrasts showing the differences in effects between native and non-native species for the covariates in Model 3. None of the contrasts from Model 3 were significant.

Model 4. The fourth mixed model included latitude only (as well as a random effect for study) and indicated a marginally significant effect of latitude in aquatic temperature studies.

Table S5.1. Parameter value estimates for the effect of latitude from Model 4, as well as contrasts between native and non-native species (the marginally significant contrast for effect of latitude is shown in bold).

System	Origin	Driver	parameter	mean	sd	val2.5pc	val97.5pc
Aquatic	Native	Temperature	beta.lat[1,1,1]	-0.1276	0.1177	-0.4065	0.04609
Aquatic	Native	CO2	beta.lat[1,1,2]	0.2541	0.1682	-0.0721	0.5876
Aquatic	Native	Plus Precip	beta.lat[1,1,3]	-0.0776	10.05	-20.06	19.57
Aquatic	Native	Minus Precip	beta.lat[1,1,4]	0.133	10.01	-19.58	19.87
Aquatic	Non-native	Temperature	beta.lat[1,2,1]	0.02346	0.07209	-0.1052	0.1989
Aquatic	Non-native	CO2	beta.lat[1,2,2]	0.2847	0.1676	-0.04197	0.6148
Aquatic	Non-native	Plus Precip	beta.lat[1,2,3]	-0.05197	10.04	-19.52	19.78
Aquatic	Non-native	Minus Precip	beta.lat[1,2,4]	0.104	9.834	-19.03	19.57
Terrestrial	Native	Temperature	beta.lat[2,1,1]	0.02218	0.07934	-0.1308	0.1878
Terrestrial	Native	CO2	beta.lat[2,1,2]	0.09549	0.135	-0.1694	0.3622
Terrestrial	Native	Plus Precip	beta.lat[2,1,3]	0.01722	0.1587	-0.2962	0.3271
Terrestrial	Native	Minus Precip	beta.lat[2,1,4]	0.05129	0.1279	-0.1983	0.2909
Terrestrial	Non-native	Temperature	beta.lat[2,2,1]	0.02884	0.07481	-0.109	0.1822
Terrestrial	Non-native	CO2	beta.lat[2,2,2]	0.1762	0.1366	-0.09007	0.4474
Terrestrial	Non-native	Plus Precip	beta.lat[2,2,3]	-0.01521	0.1564	-0.3197	0.2944
Terrestrial	Non-native	Minus Precip	beta.lat[2,2,4]	-0.002346	0.0933	-0.188	0.1804
Contrast Na	ative – Non	-native effects					
Aquatic		Temperature	cont.lat[1,1]	-0.1511	0.1243	-0.4282	0.05067 *
Aquatic		CO2	cont.lat[1,2]	-0.03065	0.1641	-0.3531	0.294
Aquatic		Plus Precip	cont.lat[1,3]	-0.02563	14.12	-27.77	27.54
Aquatic		Minus Precip	cont.lat[1,4]	0.02903	13.93	-27.49	27.15
Terrestrial		Temperature	cont.lat[2,1]	-0.006657	0.08758	-0.1791	0.1677
Terrestrial		CO2	cont.lat[2,2]	-0.08069	0.08064	-0.2403	0.0772
Terrestrial		Plus Precip	cont.lat[2,3]	0.03243	0.1334	-0.2299	0.2925
Terrestrial		Minus Precip	cont.lat[2,4]	0.05364	0.1169	-0.1805	0.2816

	Negative res	oonses	Positive responses			
Parameter	Detrimental effects of	climate change	Beneficial effects o	f climate change		
i urumeter	Native	Non-native	Native	Non-native		
Maximum effect size (a)		i ton nutive	1 uur ve	Ton nutive		
Overall	0 73(0 32)	1.55(0.41)	0.91(0.4)	1.28(0.42)		
o verun	03141	0.6.2.41	0.37205	0 55 2 25		
Terrestrial	1.05(0.32)	1.67(0.49)	1 11(0 38)	1.46(0.41)		
	0 58 1 78	0.84.2.77	0 54 1 98	0 79 2 46		
Aquatic	0.92(0.24)	1 97(0 58)	1 33(0 456)	1 76(0 55)		
Aquatic	0.52(0.24)	0.96.3.26	0 58 2 87	0.88.3.14		
Terrestrial-temperature	1.15(0.42)	1 86(0 69)	1 68(0 23)	218(0.64)		
Terrestriar temperature	0.59216	0.81.3.51	1 27 2 2	1 18 3 63		
Terrestrial-precipitation	0.99(0.24)	1.57(0.48)	0.69(0.15)	1.10,5.05		
rentestial-precipitation	0.63 1 59	0.81.2.7	05111	0.8.1.34		
Terrestrial-CO ₂	1 13(0 39)	1 64(0 56)	0.3,1.11 0.82(0.33)	1.5(0.61)		
	0.6212	0.71.2.92	0.44 21 68	0.663.05		
Aquatic-temperature	0.0,2.12	2.34(0.63)	1 25(0 56)	211(0.64)		
Aquatic temperature	0.7 1 11	1 18 3 69	0.47.2.7	1 07 3 58		
Aquatic - CO.	0.9(0.26)	1.97(0.6)	1.46(0.65)	1.07, 5.50 1.84(0.62)		
Aquatic - CO ₂	0.59.1.52	0.89.3.32	0.57312	0.84 3 39		
Half saturation constant (h)	0.37,1.32	0.07,5.52	0.57,5.12	0.04,5.57		
Overall	0.06(0.127)	0.04(0.17)	0.07(0.26)	0.16(0.47)		
0 ver un	0.000008.0.53	0.04(0.17)	0.000(0.20)	0.10(0.47)		
Terrestrial	0.51(0.32)	1 21(0.6)	0.00001.0.72 0.34(0.29)	0.000007,1.55 0.81(0.52)		
Terrestria	0.05.1.28	0.18.2.5	0.02.1.16	0.08 21 96		
Aquatic	0.03, 1.20 0.27(0.29)	1 5(0 55)	0.02,1.10 0.92(0.64)	1.17(0.51)		
Aquatic	0.27(0.29)	0 39 2 62	(0.1.2(0.04))	0.25.2.23		
Terrestrial-temperature	0.000, 1.0	0.59,2.02 0.69(0.44)	(0.1, 2.04) 0 2(0 08)	0.23, 2.23 0.75(0.38)		
Terrestrial-temperature	0.04(0.27)	0.05(0.44)	0.2(0.00)	0 2 1 64		
Terrestrial-precipitation	0.58(0.39)	1.27(0.71)	0.00,0.4 0.14(0.19)	0.09(0.09)		
refrestrat-precipitation	0.07 1 57	0.22301	0.04(0.17) 0.003.0.7	0.002.0.33		
Terrestrial- CO ₂	0.53(0.35)	1 35(0 69)	0.003,0.7 0.44(0.4)	1.007(0.63)		
	0.05 1.343	0.26.3.15	0.02.1.51	0 15 2 63		
Aquatic-temperature	0.03,1.343	1 4(0 54)	1.12(0.77)	1.07(0.45)		
inquatie temperature	0.0012(0.02)	0 4 2 54	0 15 3 11	0.34.2.1		
Aquatic - CO2	0.17(0.25)	1 64(0 68)	1 02(0 76)	1 32(0 66)		
	0.008.0.81	0 39 3 16	0.07.3.03	0.26.2.96		
Variances	0.000,0.01	0.59,5.10	0.07,5.05	0.20,2.90		
$\frac{v u n u n c c s}{n^2}$	0.14((1.72)	0.130	1.61)		
"1	0.12	0.17	0.11	0.15		
σ^2 terrestrial	0.02(.01)	0.03(0.02)	0 2(0 11)	0 14(0 05)		
	0.005 1 5	0.006 3 36	0.04 5 47	0.02 7 84		
σ^2 aquatic	0.02(0.01)	0.05(0.02)	0.03(0.02)	0.04(0.02)		
Sa uquutt	0.004 2 38	0.007 11 4	0.006 7 94	0.007 14 73		
σ^2 terrestrial	0.02(0.017)	0.06(0.02)	0.02(0.01)	0.1(0.04)		
S ₀ terrestrial	0.02(0.017)	0.008 5 33	0 004 2 24	0.01.7.63		
σ_{b}^{2} aquatic	0.02(0.016)	0.04(0.02)	0.04(0.02)	0.04(0.02)		

Appendix S6 Posterior mean (SD) and 95% CI parameter values for the hierarchical analyses. Cases where native and non-native species are significantly different (i.e. non-overlapping 95% CIs) are indicated in **bold**.

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σ_{al}^{2}	0.004,4.88 0.11(0.06)	0.006,11.93 0.06(0.03)	0.006,11.93 0.11(0.04)	0.006,10.07 0.07(0.03)
	0.02,2.05	0.01,3.55	0.01,3.95	0.01,4
σ_{bl}^{2}	1.03(0.1)	0.15(0.12)	0.97(0.09)	0.97(0.09)
	0.23,11	0.24,10	0.22,9.9	0.22,9.8

Appendix S7 Results of hierarchical analyses by system and climate driver. Observed (symbols) and predicted effect size (mean middle lines, and 95% PI lower and upper lines) as a function of magnitude of treatment are given for native (black circles and solid lines) and non-native species (gray triangles and dashed lines). Analyses were conducted separately for species responding negatively (left panels) and positively (right panels) to climatic changes.





